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Abstract

Transformation of organic microcontaminants (OMCs) during wastewater treatments results in the generation of transformation products (TPs), which can be more persistent than parent compounds. Due to reuse of reclaimed wastewater (RWW) for crop irrigation, OMCs and TPs are released in soils being capable to translocate to crops. Furthermore, OMCs are also susceptible to transformation once they reach the soil or crops. The recalcitrant antiepileptic carbamazepine (CBZ) and some of its frequently reported TPs have been found in agricultural systems. However, there is no knowledge about the fate in reuse practices of multiple CBZ TPs that can be formed during wastewater treatment processes. For the first time, this work presents a study of the behavior of CBZ TPs generated after a conventional Ultraviolet-C (UVC) treatment in an agricultural environment. The UVC-treated water was used for the irrigation of lettuces grown under controlled conditions. The latter was compared to the fate of TPs generated in the peat and plant by irrigation with non-treated water containing CBZ. A suspect screening strategy was developed to identify the TPs using liquid chromatography coupled to quadrupole-time-of-flight (LC-QTOF-MS). The results revealed the presence of 24 TPs, 22 in UVC-treated water, 11 in peat and 9 in lettuce leaves. 4 of the TPs identified in peat (iminostilbene, TP 271B, TP 285A-B); and 3 in leaves (10-11 dihydrocarbamazepine, TP 271A-B) were not previously reported in soils or edible parts of crops, respectively. Comparing the TPs found in peat and lettuces derived from both irrigation conditions, no significant differences regarding TPs formation or occurrence were observed. UVC treatment did not contribute to the formation of different TPs than those generated by transformation or metabolism of CBZ in peat or plant material. This research improves the current knowledge on the fate of CBZ TPs in agricultural systems as a consequence of reuse practices.

Keywords: Carbamazepine, transformation products, LC-QTOF-MS, wastewater reuse, suspect screening

1. Introduction

Nowadays, standard treatment processes applied in wastewater treatment plants (WWTPs) do not remove efficiently a large variety of organic microcontaminants (OMCs) as pharmaceuticals, personal care products or pesticides (Campos-Mañas et al., 2017). With OMCs, several recalcitrant transformation products (TPs), generated during the treatments, are continuously discharged in WWTP effluents (Schollée et al., 2015). As agricultural practices demand a large amount of water, reuse of reclaimed wastewater (RWW) has become a common practice in many dry areas to deal with water shortages. Consequently, OMCs and TPs have been reported in agricultural soils at concentrations up to μg g^{-1} (Chen et al., 2011; Christou et al., 2017; Kinney et al., 2006; Koba et al., 2016). Due to their physical-chemical properties, some of these compounds have the potential to be uptaken via plant roots (Wu et al., 2015). Once compounds have entered the plant, a subsequent translocation toward other parts of plants, including the edible part of crops, can take place resulting in the possible introduction of undesirable substances into the food chain. Although the number of studies dedicated to soil accumulation and plant uptake of OMCs is steadily increasing in recent years (Carter et al., 2018; Larivière et al., 2017; Martínez-Piernas et al., 2018a), little information is available regarding TPs behavior in soil/plant systems. These TPs often present similar or even greater concentration levels than their parent compounds in WWTP effluents (Bahlmann et al., 2014). Additionally, they can be also generated in soils from biotic/abiotic transformations and in crops as a consequence of the metabolism of plants (Huynh et al., 2018; Riemenschneider et al., 2017). Considering that some TPs have analogous or even more severe biological activity than parent compounds (Brezina et al., 2017), their fate and ecotoxicological and human health risks merit further research.

Generally, TPs show very diverse physical-chemical properties due to their different structures. For this reason, broad spectrum extraction methodologies are required to obtain efficient recoveries in a wide range of compounds . QuEChERS (Quick, Easy,Cheap, Effective, Rugged and Safe) and pressurized liquid extraction (PLE) have demonstrated to be good alternatives even in these complex environmental matrices (Martínez-Piernas et al., 2018b) (Jelić et al., 2009). Besides, the application of screening methodologies accomplished by liquid chromatography coupled to high resolution mass

spectrometry (HRMS), have undoubtedly improved the identification of unexpected or not previously validated compounds by the application of non-target and suspect screening approaches (Martínez-Piernas et al., 2018a).

Carbamazepine (CBZ) is one of the most frequently detected OMCs in WWTP effluents due to its recurrent prescription for neuropsychiatric disorders (Ambrósio et al., 2002) and its low removal by the application of standard wastewater treatment processes (Zhang et al., 2008). Because of its persistence and ubiquitous occurrence, it has been proposed as an appropriate indicator for the evaluation of anthropogenic impact on the aquatic environment (Kinney et al., 2008). According to the criteria established by Council Directive 92/32/EEC, CBZ has been classified as potentially harmful compound for aquatic organisms (Fent K., 2008). In addition, the formation of several of its TPs has been reported by the application of different wastewater treatments, which are presented in Table S1. CBZ undergoes transformation to various aldehydes, ketones and hydroxylated derivatives and known ecotoxic compounds as acridine and acridone (Donner et al., 2013). However, information about presence and fate of CBZ TPs in soils and crops is still scarce. Riemenschneider et al. (Riemenschneider et al., 2017) investigated the formation and translocation of CBZ TPs through the different parts of tomato plants irrigated with a spiked solution of CBZ under hydroponic conditions. Regarding soils, Koba et al. (Koba et al., 2016) evaluated the stability of CBZ in different soils, identifying in samples up to three TPs after an incubation process with CBZ. Nevertheless, to our knowledge, no data is available about the fate of CBZ TPs produced after standard tertiary treatments in agricultural systems. Regarding tertiary treatments, advanced oxidation processes (AOPs), which are characterized by the formation of powerful oxidizing species, have been proved to be effective in the degradation of organic contaminants (Malato et al., 2009). Among available AOPs, ultraviolet treatment is one of the most extended processes for drinking and wastewater purification and, in particular, degradation of CBZ by UV and UV-based AOP has been widely reported in literature (Dai et al., 2012; Deng et al., 2013; Ghasemian et al., 2017).

The objective of this study was to increase the current knowledge on the fate of CBZ TPs in an agricultural system based on a lettuce crop grown in peat under controlled conditions. The presence and fate of TPs in these two commodities were compared

considering two different TP sources: i) TPs formed after conventional Ultraviolet-C (UVC) treatment applied to the irrigation water, and ii) TPs formed in the soil-plant system by the transformation of CBZ introduced by theirrigation water. The identification of CBZ TPs was carried out by the application of a suspect screening strategy by LC-QTOF-MS, which included up to 47 TPs commonly found after different decontamination or biological processes.

2. Materials and methods

2.1 Chemicals

Carbamazepine (CBZ), carbamazepine 10,11-epoxide (EPOX), acridone (ACRO), acridine (ACRI), oxcarbazepine (OX) and carbamazepine-d_{10} (CBZ-d_{10}) analytical standards (purity grade ≥ 98%) were purchased from Sigma Aldrich (Steinheim, Germany). Iminostilbene, 9-acridinecarboxylic acid, 9-acridine-carboxaldehyde and 10-11 dihydrocarbamazepine (all purity ≥ 98%) identified by suspect screening analysis and acquired for confirmatory purposes were also purchased from Sigma Aldrich. LC-MS grade acetonitrile (MeCN), methanol (MeOH), water, formic acid and acetic acid were purchased from Sigma Aldrich. For QuEChERS, magnesium sulfate ($MgSO_4$), sodium acetate (NaOAc), octadecyl silica (C18) and primary-secondary amine (PSA) were purchased from Sigma Aldrich. Hydromatrix was provided by Thermo Fisher Scientific (Waltham, USA).

Stock standard solutions were prepared in MeOH at a concentration of 1000 mg L^{-1}. A mixed working solution containing all standards was prepared at 10 mg L^{-1} in MeOH by proper dilution of each stock standard solution. All solutions were prepared in amber glass vials and stored at -20 °C. CBZ-d_{10} was used as extraction quality control check.

2.2 Experimental set-up

2.2.1 Experimental lettuce cultivation

Seeds of lettuce (*Lactuca sativa*) obtained from a local provider were cultivated under controlled conditions of temperature and humidity in an experimental greenhouse described by Martínez-Piernas et al., 2018b. 90 propylene pots (9 × 9 × 10 cm) were filled with sterilized peat (autoclaved using autoclave-bags at 121 ºC during 15 min in

batches of 5 kg of peat). The peat was a mixture of blond peat, black peat, coconut fibers and perlite containing N, P, and K in a ratio (w/v) of 13–14–13 g L^{-1}, respectively, pH 7, organic matter dry matter ratio of 80%, apparent density of 0.38 kg L^{-1} and 120 mS m^{-1} of conductivity, according to the manufacturer. The growing crops was not done under sterile conditions. The growing period was conducted from May to July 2016, a total of 10 weeks. Three experimental conditions (30 pots each) were performed separately to avoid any cross-contamination: a) control samples irrigated with synthetic water (SW); b) samples irrigated with SW spiked with 1 mg/L of CBZ (SW+CBZ); and c) samples irrigated with SW spiked with 1 mg L^{-1} of CBZ and treated by UVC (SW+CBZ+UVC). Pots were irrigated every two days. The experimental setup for the three cultivations of lettuce crops is shown Figure S1. The sampling strategy was designed to evaluate potential presence and accumulation of CBZ and metabolites/TPs in peat and lettuce leaves. A total of five sampling events occurred. Samples were taken every two weeks from the second week of growth until the tenth week (harvest). In each sampling event, ten pots randomly selected were taken (leaves and peat) and combined to form a homogenized composite sample which was extracted per triplicate. The final size of lettuce leaves was 15 cm in the last sampling event.

2.2.2 Irrigation water

SW was prepared following the recipe published in (American Public Health Association, American Water Works Association, 2012) under the "standard moderately-hard freshwater" nomenclature, based on the characteristics of groundwater in Almería province (Spain). For the irrigation of crops with CBZ, SW was spiked with the appropriate amount of pure CBZ standard to reach a final concentration of 1 mg L^{-1}. Before each irrigation event, a fresh solution of CBZ was prepared to avoid the possible formation of undesirable TPs. For the irrigation tests with treated-CBZ, UVC treatments were carried out in a pilot plant previously described by Miralles-Cuevas et al., 2017. Briefly, it consists of three independent low-pressure UVC lamps (254 nm peak wavelengths, 230 W and 40 mJ cm^{-2} of UV dose or fluence) serially connected to holding tank. The volume of each lamp-camera is 5 L. In this work, the system was operated with one UVC lamp in recirculating batch mode at 30 L/min of flow. The tank was filled with 80 L of SW and spiked with CBZ (1 mg L^{-1}). After 10 min of mixing in the dark, a Time 0 was taken out and the UVC lamp was switched on. From this time,

samples were taken every 2 min during the first 20 min, and every 5 min till the end of the treatment (60 min total exposure time). The treated water (ca. 60 L) was stored at 4ºC and used for crop irrigation during one week. The same procedure was repeated weekly during the irrigation period (a total of 10 weeks) in order to use fresh-batches and avoid possible fluctuations of TPs during storage. Irrigation events occurred every two-three days depending on plant water demand, with 50 mL of water/pots. All water batches were analyzed by LC-QTOF-MS before irrigation to verify the absence of any compound in control water (SW), the absence of TPs in water spiked with CBZ (SW+CBZ) and possible fluctuations in the formation of TPs in treated water (SW+CBZ+UVC).

2.3 Sample preparation

2.3.1 Lettuce extraction

Leaves of lettuce samples were washed with tap water, chopped and stored in the dark at -20˚C until their analysis. Samples were extracted by a QuEChERS-based extraction method including a dispersive solid-phase extraction (d-SPE) clean-up step (Martínez-Piernas et al., 2018b). Briefly, a representative aliquot of 10 g of previously homogenized sample was weighed in a 50 mL PTFE centrifuge tube. 10 mL of MeCN at 1% of acetic acid and 50 µL of CBZ-d_{10} (400 µg L^{-1}), used as internal quality control, were added and the tube was shaken for 5 min. After that, 6 g of $MgSO_4$ and 1.5 g of NaOAc were added and the tube was vigorously shaken for 5 min and centrifuged at 3500 rpm (2054 *g*) for 5 min. Then, a 5 mL aliquot of the organic layer was transferred to a 15 mL centrifuge tube containing 125 mg of PSA, 125 mg of C18 and 750 mg of anhydrous $MgSO_4$. The tube was then shaken for 30 s in a Vortex and centrifuged again (3500 rpm, 5 min). After that, the extracts (4 mL) were transferred to screw cap vials and 10 µL of MeCN 1% formic acid per mL of extract were added. Finally, an aliquot of 150 µL of the extract was evaporated until dryness and reconstituted with the same volume of MeCN:H_2O (10:90, *v/v*) before the injection in the LC-QTOF-MS/MS system.

2.3.2 Peat extraction

Peat samples were homogenized, freeze dried and finally grinded using a Mixer Mill MM 301 equipped with two cells of 35mL made of ZrO_2. Samples were extracted by

PLE following the protocol described in (Jelić et al., 2009) using an ASE 300 accelerated solvent extractor followed by a solid-phase extraction (SPE) clean-up step. 1 g of homogeneous freeze-dried peat was placed in a stainless steel extraction cell of 11 mL, which was filled with hydromatrix. The extraction solvent consisted of a mixture of MeOH:H_2O (1:2, *v/v*). Optimized PLE parameters chosen were: a temperature 100 ºC, a preheating period of 5 min, a total of 3 static cycles (5 min each), and total flush volume of 100% of the cell with 60 s of nitrogen purge. PLE extract (about 40 mL) was diluted in 500 ml of H_2O and cleaned-up by SPE using Oasis HLB cartridges (200 mg, 6 mL). The cartridges were conditioned with 5 mL of MeOH followed by 5 mL of H_2O at neutral pH. The elution of compounds was carried out with 8 mL of MeOH. Then, SPE extracts were evaporated under nitrogen stream and reconstituted in 1 mL of MeCN:H_2O (10:90, *v/v*) before LC-QTOF-MS/MS injection.

2.4 Analysis by liquid chromatography tandem mass spectrometry

Chromatographic separation was carried out using a HPLC 1260 Infinity (Agilent Technologies, Palo Alto, CA, USA) system provided with an Eclipse C18 (4.6 x 150 mm, 5µm particle size) column (Agilent Technologies). The mobile phases were 0.1% formic acid in water (solvent A) and pure MeCN (solvent B). The injection volume was 20 µL and the flow rate was 0.5 mL min^{-1}. The initial proportion of solvent B was 10%, which was kept constant for 2 min, increased to 100% within 38 min, kept constant for 10 min and reduced to 10% in 0.1 min. The post-run equilibration time was 15 min.

For HRMS, a TripleTOF® 5600+ System (Sciex, Foster City, CA, USA) equipped with a dual source was used. ESI interface was employed for sample injection and the atmospheric-pressure chemical ionization interface (APCI) for calibrant delivery. The ESI source was operated in positive mode. The parameters applied were 60 psi of gas 1 and 2,; 30 psi of curtain gas,; an ionspray voltage of 4500 V; a declustering potential of 80 V and a temperature of 575 ºC. Nitrogen was used as nebulizer, curtain and collision gas. The acquisition method consisted in a full-scan survey (TOF-MS) followed by four TOF-MS/MS scans carried out by Information Dependent Acquisition (IDA) of the four more intense ions in each TOF-MS scan. Scanned mass range was from 50 to 1000 *m/z*, either in TOF-MS (resolving power of 30000) or TOF-MS/MS experiments. An accumulation time of 250 ms was applied in TOF-MS and 100 ms for IDA scan. IDA

criteria considered dynamic background subtraction. Collision energy of 30 eV with a ± 15 eV spread was used in MS/MS fragmentation. Data acquisition was carried out by Analyst TF 1.5, and data processing by PeakView™ 2.2 and MasterView 1.1.

2.5 Suspect screening strategy

A suspect list including 47 possible transformation and biotransformation products of CBZ was built according to the previously reported TPs in literature generated by diverse decontamination wastewater treatments and biological processes (Table S1). As first step of data processing, a reduction of the number of peaks for a reliable identification was carried out by applying a peak intensity threshold ≥ 1000 cps, a S/N ratio ≥ 10 and the absence of the mass in the control sample (blank matrix). After that, the criteria adopted for a tentative identification was a mass accuracy error ≤ 5 ppm of the precursor ion and an isotope ratio difference ≤ 10%. The MS/MS information was compared with spectra reported in literature, MassBank ("MassBank Database," n.d.) and ChemSpider ("ChemSpider Database," n.d.) databases; for which a minimum score of 80% and presence of at least two fragments with an accurate mass error ≤ 5 ppm were considered acceptable. Final confirmation of tentative identified compounds was adopted when the retention time (Rt) of the standard in matrix differed less than ± 0.1 min and the MS/MS spectra matched.

The TPs tentatively identified were grouped according to the confidence levels proposed by Schymanski et al. (Schymanski et al., 2014). Level 4 included TPs for which enough MS/MS fragmentation information was not acquired and, consequently, no structure could be suggested. Level 3 was adopted for those compounds whose MS/MS information matched with literature or libraries, but different structures could be proposed. In Level 2 were accommodated compounds with enough MS/MS and experimental context data to propose a unique probable structure. Finally, Level 1 was considered for TPs confirmed by the unequivocal information of Rt and MS/MS fragmentation of the purchased analytical standard.

2.6 Methods validation

Both QuEChERS-based and PLE+SPE, procedures applied in this study for the extraction of CBZ and TPs in plant material and peat were validated for a set of 5

compounds: CBZ, EPOX, ACRI, ACRO and OX. The validation was carried out in terms of linearity, limits of quantification (LOQs), trueness (recoveries) and precision (relative standard deviations, RSD). For validation purposes, control samples of lettuce and peat were used as blanks.

Linearity was studied by spiking matrix blank extracts at 11 different concentrations ranging from 0.1 to 200 ng g^{-1}. Adequate determination coefficients (R^2) were considered acceptable when $R^2 \geq 0.990$. Recoveries and precision (n=3) were evaluated by spiking blank samples (20 ng g^{-1} in peat and 10 ng g^{-1} in lettuce). LOQs were experimentally calculated as the lowest concentration level spiked in blank matrix extract which fulfill the requirements of analyte confirmation. In Table S2 is compiled the information of both validation methodologies.

3. Results and Discussion

3.1 Identification of CBZ TPs by suspect screening

Following the suspect screening approach described above (Section 2.5.), a total of 24 TPs out of the 47 included in the suspect list were tentatively identified in some of the analyzed samples (irrigation water, peat or lettuces). The list of candidates and chromatographic and identification information is presented in Table 1.

One of the main difficulties regarding TPs identification is the differentiation between isomers or compounds with closely related structures, which can show the same accurate mass and elemental composition and even very similar MS/MS fragmentation due to their related structures. This can lead to flimsy or erroneous tentative identifications, even when an adequate chromatographic separation is carried out. This is the case of the 12 TPs classified in Level 3, for which varied structures could be proposed in each case.

An example regarding difficulties on appropriate identification of TPs in absence of standards was the allocation of structures for TPs 253A-D, with extraction mass *m/z* 253.0971 for the $[M+H]^+$. The elemental composition, $C_{15}H_{12}N_2O_2$, corresponded to the addition of one oxygen atom to the structure of CBZ and all of them presented the same fragmentation pattern. Fig. 1 shows the extracted ion chromatogram (XIC: 253.0971 m/z) and MS/MS spectra of TPs 253B-D in an irrigation water sample, where the

similarity of the MS/MS spectra for the three compounds can be appreciated. Among the structures found according to the proposed formula (Table S1), the fragmentation pattern matched well with the spectra of the isomers typically produced by the monohydroxylation of CBZ (OH-CBZ) at different positions, as proposed by several authors (Ahmed and Chiron, 2014; Brezina et al., 2017; Hübner et al., 2014; Jelic et al., 2013; Li et al., 2013; Liu et al., 2016; Zhang et al., 2015; Zhu et al., 2016). Up to three isomers have been reported with hydroxylation at positions 2, 3 and 10. However, analysis of the available analytical standards allowed confirmation (Level 1) of TP 253C as EPOX (Ahmed and Chiron, 2014; Hübner et al., 2014; Li et al., 2013; Liu et al., 2016; Zhang et al., 2015; Zhu et al., 2016) and TP 253D as OX (Brezina et al., 2017), as it is shown in Fig. 1 . In this case, only the Rt comparison with the analytical standard allowed to distinguish both compounds. 253B was then tentatively assigned as a monohydroxy derivative, but the Rt and spectral information available was not enough to clarify the position of the hydroxylation in the ring. Therefore, TP 253B was kept in Level 3. TP 253A, also presented the same molecular formula and isotopic profile and could be tentatively proposed as a second OH-CBZ isomer, but due to its low intensity, not enough product ions with significant intensities and exact mass data were acquired. For this reason, it was considered in identification Level 4.

A similar situation was observed for TP 269 (*m/z* 269.0920, $C_{15}H_{12}N_2O_3$). Up to five peaks, TPs 269 A-E were detected. TP 269B remained in Level 4 by the same reason already exposed for TP 253A. The other four peaks presented similar characteristic fragments at *m/z* 251.0815, *m/z* 208.0757 and *m/z* 180.0808. The elemental composition, with two additional oxygen atoms with respect to CBZ, was in accordance with the formation of dihydroxy-CBZ derivatives proposed by Hübner et al (Hübner et al., 2014), and TP 269A and 269C were tentatively proposed as dihydroxylated isomers. However, other structures with alike fragments have also been reported in literature by Ahmed and Chiron (2014), Jelic et al., (2013) and Zhu et al., 2016(see Table S1). The hydroxylation of OX intermediate proposed by Jelic et al. (Jelic et al., 2013) match with the mass spectrum of TP 269D (Fig. S2A), which shows the characteristic fragment at *m/z* 196.0757, corresponding to the formation of hydroxyl acridine ($C_{13}H_9NO$). However, some differences in the spectrum reported for 11-OH-OX (Jelic et al., 2013) can be explained by the different position of the OH group, as it is proposed in Fig. S2. On the other hand, the absence of the diagnostic fragment at *m/z* 196.0757 in the mass

spectrum of TP 269E (Fig. S2B) suggested a different structure for this compound. The alternative proposed by Ahmed and Chiron corresponds to the hydroxylation of 9-formylacridine-10(9H)-carboxamide (Ahmed and Chiron, 2014), which can be plausible and supported by the successive losses of CHNO (*m/z* 226.0863), H_2O (*m/z* 208.0757) and CO (*m/z* 180.0808) observed in the mass spectra (Fig. S2B). A subsequent hydroxylation of TP 269E would be consistent with the formation of dihydroxy derivatives also identified as TP 285A and B (Fig. S3).

Identification of TPs 208A and 208B (*m/z* 208.0756; $C_{14}H_9NO$), could not be carried out. Two structures could fit with this *m/z* corresponding to 9-acridine-carboxaldehyde (Hübner et al., 2014; Liu et al., 2016; Seiwert et al., 2015; Zhang et al., 2015; Zhu et al., 2016), which was discarded by the analysis of the analytical standard, and the human metabolite CBZ iminoquinone (Brezina et al., 2017), which shared a characteristic product ion at *m/z* 152.0495 ($C_{11}H_5N$), with TP 208B. However, this evidence was too weak for the allocation of the CBZ iminoquinone structure and both compounds remained in Level 4.

TPs 224A (Rt 4.7 min) and 224B (Rt 22.7 min) were detected at *m/z* 224.0706 ($C_{14}H_9NO_2$). Both presented the same MS/MS fragments. Some authors have associated this formula to varied structures (Brezina et al., 2017; Hübner et al., 2014; Jelic et al., 2013; Li et al., 2013; Riemenschneider et al., 2017), describing common product ions in many cases (Table S1). One of the most plausible ones for TP 224A, due to its polar chromatographic behavior, was 9-acridinecarboxylic acid, which was confirmed by the analytical standard. The retention time behavior of TP 224B could match with varied structures. However, based on the MS/MS spectrum, the compound was tentatively proposed as acridone-N-carbaldehyde, according with Li et al. (Li et al., 2013) in a study about identification and kinetic of metabolites of CBZ in soil. The similarity with the fragmentation pattern of acridone after the loss of -CO (*m/z* 196.0750; $C_{13}H_9NO$) supported the proposal of this structure at Level 3.

In the case of TPs 267A and 267B (*m/z* 267.0764; $C_{15}H_{10}N_2O_3$), only the first one was tentatively proposed as 11-keto oxcarbazepine, based on the MS/MS fragmentation pattern reported by Jelic et al. (2013) and Koba et al. (2016). Many other structures have been reported for TPs with *m/z* 267.0764, however not enough evidences have

been found that support a structure assignation. Consequently, TP 267B was not assigned (Level 4).

TPs 271A and B, also presented very similar fragmentation, which matched well with that proposed by Jelic et al. (Jelic et al., 2013) and Hübner et al. (Hübner et al., 2014) for 10,11-dihydro-10,11-dihydroxycarbamazepine. Li et al. (Li et al., 2013) also confirmed this compound by comparing with an authentic standard and reported the presence of the cis and trans stereoisomers, which could correspond with the two peaks observed. The assignation of this structure was also reinforced with the identification of TP 287 (*m/z* 287.1026; $C_{15}H_{14}N_2O_4$), which could correspond with a further hydroxylation of the benzene ring in TP 271.

Despite of the general absence of TPs MS/MS spectra in spectral libraries and databases, attributable to the scarce availability of commercial analytical standards, some of the TPs under study could be identified by this way. This was the case of TP 194 (*m/z* 194.0964, $C_{14}H_{11}N$), which matched with iminostilbene structure (94% score) in MassBank, or TP 239 (*m/z* 239.1179, $C_{15}H_{14}N_2O$), which revealed a 90% of spectral score match with 10,11-dihydrocarbamazepine in ChemSpider database. Both were confirmed by subsequent standard acquisition and analysis.

Although spectral and context evidences pointed out a tentative structure proposal for most of the compounds investigated, a definite confirmation by analytical standard (Level 1) was only obtained for 7 TPs namely ACRI, ACRO, EPOX, OX, TP 194 (iminostilbene), TP 224A (9-acridinecarboxylic acid) and TP 239 (10-11 Dihydrocarbamazepine).

3.2 Identification of CBZ TPs in irrigation water, peat and lettuce leaves

It is already known that CBZ is a recalcitrant compound whose removal is not efficient by conventional treatments, leading to its constant detection in WWTP effluents (Campos-Mañas et al., 2017). In this study, the removal of CBZ in water after UVC treatment described in the experimental section was about 20 %, as it is shown in Fig. S4. However, despite its persistence, a total of 22 CBZ TPs were identified in the treated water (SW+CBZ+UVC), which was used in the irrigation assays (see Table 2).

In general, peak areas were comparable in each irrigation batch, showing a repetitive pattern of TPs formation. The most abundant TP found in treated water was EPOX followed by TP 253B (OH-CBZ), TP 194 (iminostilbene) and ACRI, which were detected from the second minute of treatment (Fig. S5). Overall, TP abundances increased with treatment time due to the persistence of CBZ. Alternatively, no TPs were detected in the irrigation water containing CBZ, which had not undergone any treatment.

For peat and lettuces irrigated with SW, neither CBZ nor any of its TPs were observed. Regarding samples irrigated with SW+CBZ and SW+CBZ+UVC, the results showed that almost identical TPs were detected in both irrigation experiments. As can be seen in Table 2, 10 TPs were identified in peat irrigated with untreated water containing CBZ and 11 in peat irrigated with the treated water, while 9 TPs were found in lettuces regardless of the water used for irrigation. These data suggest that UVC treatment did not contribute to the presence of different TPs with respect to those formed by the transformation of CBZ in peat or lettuce. TPs formation was also possible by the only presence of CBZ in irrigation water. This is in agreement with the results found by Riemenschneider et al. (Riemenschneider et al., 2017) for tomato plants cultivated under hydroponic conditions irrigated with a nutrient solution containing only CBZ.

In regard to the abundances of the identified TPs, these were higher, in general, in peats irrigated with UVC-treated water since the treatment promotes the TPs formation. Fig. 2 shows the evolution on the abundances of the CBZ TPs detected in lettuce and peat samples during the plant growth. Higher differences were observed in TPs 271C and 285B. This pattern was also observed when concentrations of EPOX, ACRI and ACRO were quantitatively evaluated. As shown in Table 3, slightly higher TP concentration values were obtained from peat irrigated with treated water, although this correlation was not always observed in lettuce. For the vegetable, the CBZ TP concentrations were higher in samples irrigated with SW+CBZ (Table 3 and Fig. 2). This demonstrates the necessity to develop efficient wastewater treatments able to completely remove recalcitrant compounds as CBZ in order to prevent their plant metabolization, which may lead to the detection of TPs in edible parts of crops. TP concentrations followed the order EPOX > ACRO > ACRI in every commodity and experimental irrigation test. EPOX has been reported as the most abundant TP in soils (Koba et al., 2016) and

tomato plants (Riemenschneider et al., 2017) exposed to CBZ for long periods. In general, TPs did not show biodegradability but a clear accumulation along sampling events was detected (Table 3), highlighting the accumulation of EPOX in both type of samples and irrigation tests. This accumulation can be explained in part by the increase in the transpiration rate associated with the growth of the lettuce plant (Dodgen et al., 2015), although other factors, such as the plant's physiology, environmental conditions and TPs physicochemical properties (i.e. lipophilicity and electrical charge), can also contribute to this behavior (Christou et al., 2017).

In Table 2, it is also shown two TPs (TP 285 A and B) detected in peat samples and not in water. Therefore, it can be hypothesized that the formation of these TPs can be attributed to the metabolism of CBZ in peat since none of them was previously identified in UVC-treated irrigation water. The absence of TP 285A in peat irrigated with SW+CBZ could be attributable to a lower formation of this isomer. Concerning lettuce samples, TP 269C and 271B were found only in lettuce leaves. They could be supposedly generated by the metabolization of CBZ in plant material. Furthermore, their absence in peat samples may also be attributed to their further mineralization or degradation to other TPs in peat.

Some of the CBZ TPs investigated in this study as ACRI, ACRO, EPOX and TP 267A, have already been reported in agricultural soils and soilless cultures (Koba et al., 2016; Li et al., 2013; Martínez-Piernas et al., 2018a). As well, ACRI, ACRO, EPOX, OX, TP 224 A (9-acridinecarboxylic acid), TP 239 (10-11 dihydrocarbamazepine), TP 253 A/B, TP 271 A/B have been identified in plant tissues as roots, stems or leaves (Martínez-Piernas et al., 2018b; Riemenschneider et al., 2017, 2016), the suspect screening approach applied has allowed the tentative identification of new TPs not previously found neither in agricultural substrate or soils nor in vegetable matrices. To the authors' knowledge, this study reports for the first time the identification of TP 194 (iminostilbene), TP 271B and TPs 285A/B in an agricultural substrate as peat, as well as TP 239 and TPs 271A/B in in a lettuce crop.

One of the main challenges regarding reuse of WW for agricultural purposes is having more knowledge about the formation and occurrence of TPs more toxic than parent compounds. Some of the TPs identified in lettuce samples in this study, as ACRI and

ACRO have exhibited more toxicity, when both analytes were found mixed, than CBZ itself across multiple trophic levels (Donner et al., 2013). Besides, in a genotoxicity prediction study carried out by Brezina et al. (Brezina et al., 2017), CBZ derivates such as 9-acridinecarboxylic acid (TP 224A) showed higher toxicological relevance than CBZ. On the other hand, EPOX has potential genotoxic carcinogenicity (Houeto et al., 2012). In this outline, it is necessary not only a toxicological evaluation of parent compounds but also taking into account mixture toxicities to evaluate human health and environmental impacts derived from reuse of RWW in agriculture.

4. Conclusions

This work presents the first evaluation of the behavior of CBZ TPs formed by a conventional UVC water treatment in an agricultural system. The UVC-treated water was used to irrigate a lettuce crop grown in peat under controlled conditions. The fate of TPs in the latter was compared to the TPs generated due to CBZ degradation processes in both commodities. For TPs identification, a rapid and semi-automatic suspect screening approach was applied to peat and lettuce samples by LC-QTOF-MS. The suspect screening strategy revealed the presence of up to 11 CBZ TPs in peat and 9 in lettuce leaves, showing the potential of the suspect screening approach. No substantial differences regarding TPs formation or fate were found derived from the diverse irrigations. In any case, TPs were likely to reach the edible parts of crops, so highlighting the need for efficient wastewater treatments able to remove OMC to avoid their translocation to plant tissues. This study has contributed to a better understanding of the fate of CBZ TPs and results obtained can serve as a basis to extend the study of these TPs to field crops, grown under diverse conditions. As a general remark, more knowledge regarding OMC TPs structure and behavior must be obtained in order to fully assess the risk associated with their discharge in the environment and human consumption due to reuse practices in agriculture.

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6. References

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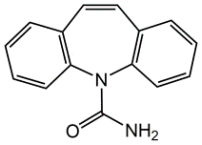
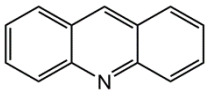
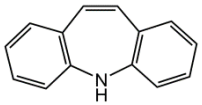
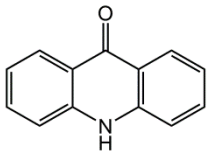
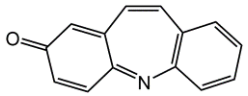
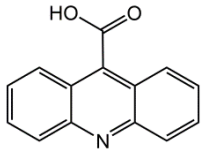
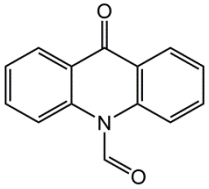
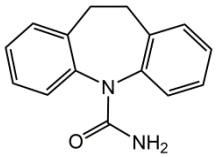
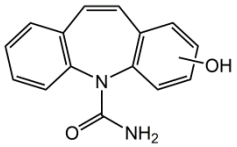
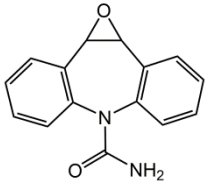
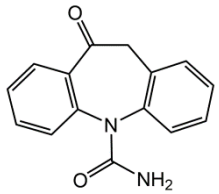
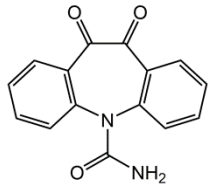
Tables

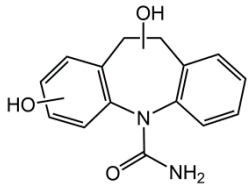
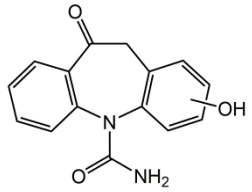
Table 1. List of CBZ TPs identified in samples, accurate mass and chromatographic information.

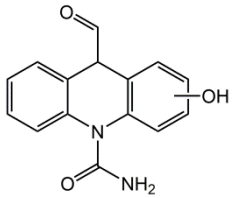
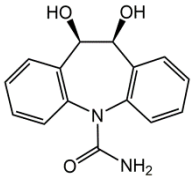
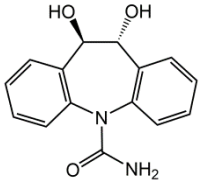
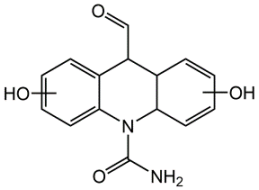
Compound	Structure	Molecular formula	$[M+H]^+$ (*m/z*)	Error (ppm)	Rt (min)	Product ion (PI)	Assigned formula	PI Error (ppm)	Identification level	Criteria	Reference
CBZ		$C_{15}H_{12}N_2O$	237.1022	-0.2	21.2	194.0964 192.0808 179.0730	$C_{14}H_{11}N$ $C_{14}H_9N$ $C_{13}H_9N$	-0.1 -2.5 0.3	L1	Standard	
TP 180 (ACRI)		$C_{13}H_9N$	180.0807	-0.8	12.0	178.0651 154.0651 153.0699	$C_{13}H_7N$ $C_{11}H_7N$ $C_{12}H_8$	2.7 -4.7 -4.9	L1	Standard	(Ahmed and Chiron, 2014; Li et al., 2013; Liu et al., 2016; Zhang et al., 2015; Zhu et al., 2016)
TP 194 (Iminostilbene)		$C_{14}H_{11}N$	194.0964	-3.7	31.8	179.0730 167.0730 152.0621	$C_{13}H_9N$ $C_{12}H_9N$ $C_{12}H_8$	-2.0 -9.9 -3.6	L1	Standard	(Liu et al., 2016)

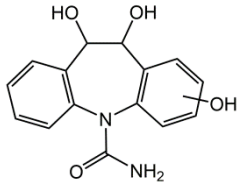
TP 196 (ACRO)		$C_{13}H_9NO$	196.0756	-1.3	19.8	178.0651 167.073 139.0542 115.0542	$C_{13}H_7N$ $C_{12}H_9N$ $C_{11}H_6$ C_9H_6	-0.7 -0.3 -1.6 2	L1	Standard	(Brezina et al., 2017; Hübner et al., 2014; Liu et al., 2016; Zhu et al., 2016)
TP 208A	No proposal	$C_{14}H_9NO$	208.0756	-0.6	26.0	190.0651 180.0808 178.0651 154.0651 153.0699	$C_{14}H_7N$ $C_{13}H_9N$ $C_{13}H_7N$ $C_{11}H_7N$ $C_{12}H_8$	0.4 4.0 -1.3 4.4 3.4	L4		(Hübner et al., 2014; Seiwert et al., 2015)
TP 208B (CBZ iminoquinone)		$C_{14}H_9NO$	208.0756	-0.6	28.7	180.0808 178.0651 152.0495	$C_{13}H_9N$ $C_{13}H_7N$ $C_{11}H_5N$	-4.9 -6.3 -4.4	L4	MS/MS spectra and RT reported	(Brezina et al., 2017; Liu et al., 2016)
TP 224A (9-acridinecarboxy-lic acid)		$C_{14}H_9NO_2$	224.0706	-1.2	4.7	196.0757 180.0808 167.0730	$C_{13}H_9NO$ $C_{13}H_9N$ $C_{12}H_9N$	4.6 2.4 1.5	L1	Standard	(Brezina et al., 2017; Hübner et al., 2014; Jelic et al., 2013; Li et al., 2013; Riemenschneider et al., 2017)

TP 224B (Acridone-N-carbaldehyde)		$C_{14}H_9NO_2$	224.0706	4.0	22.7	196.0757 180.0808 167.0730	$C_{13}H_9NO$ $C_{13}H_9N$ $C_{12}H_9N$	0.6 2.9 -2.1	L3		MS/MS spectra	(Li et al., 2013)
TP 239 (10-11 Dihydrocarbamazepine)		$C_{15}H_{14}N_2O$	239.1179	0.5	21.5	196.1121 194.0964 180.0808	$C_{14}H_{13}N$ $C_{14}H_{11}N$ $C_{13}H_9N$	-5.0 -1.7 0.7	L1		Standard	(Stein et al., 2008)
TP 253A		$C_{15}H_{12}N_2O_2$	253.0971	-3.8	13.5	No MS/MS			L4			
TP 253B (OH-CBZ)		$C_{15}H_{12}N_2O_2$	253.0971	1.4	17.3	236.0706 210.0913 208,0757 182.0964 180.0808 167.0730	$C_{15}H_9NO_2$ $C_{14}H_{11}NO$ $C_{14}H_9NO$ $C_{13}H_{11}N$ $C_{13}H_9N$ $C_{12}H_9N$	4.5 -0.7 4.0 -4.0 -6.5 -2.7	L3		MS/MS spectra reported	(Jelic et al., 2013)
TP 253C (EPOX)		$C_{15}H_{12}N_2O_2$	253.0971	0.4	18.2	236.0706 210.0913 208,0757 182.0964 180.0808 167.0730	$C_{15}H_9NO_2$ $C_{14}H_{11}NO$ $C_{14}H_9NO$ $C_{13}H_{11}N$ $C_{13}H_9N$ $C_{12}H_9N$	-6.4 -1.6 -1.9 -2.9 -0.4 0.9	L1		Standard	(Ahmed and Chiron, 2014; Hübner et al., 2014; Li et al., 2013; Liu et al., 2016; Zhang et al., 2015; Zhu et al.,

											2016)
TP 253D (OX)		$C_{15}H_{12}N_2O_2$	253.0971	0.6	18.9	236.0706 210.0913 208,0757 182.0964 180.0808 167.0730	$C_{15}H_9NO_2$ $C_{14}H_{11}NO$ $C_{14}H_9NO$ $C_{13}H_{11}N$ $C_{13}H_9N$ $C_{12}H_9N$	0.2 -1.6 0.1 -4.0 -4.3 -4.5	L1	Standard	(Brezina et al., 2017)
TP 267A (11-Keto oxcarbazepine)		$C_{15}H_{10}N_2O_3$	267.0764	1.3	15.3	239.0815 224.0706 196.0757 168.0808 212.0706	$C_{14}H_{10}N_2O_2$ $C_{14}H_9NO_2$ $C_{13}H_9NO$ $C_{12}H_9N$ $C_{13}H_9NO_2$	3.6 3.6 4.9 8.5 8.5	L3	MS/MS spectra reported	(Ahmed and Chiron, 2014; Brezina et al., 2017; Hübner et al., 2014; Jelic et al., 2013; Koba et al., 2016; Li et al., 2013; Zhu et al., 2016)
TP 267B	No proposal	$C_{15}H_{10}N_2O_3$	267.0764	0.3	18.8	224.0706 222.0550 206.0600 196.0757 167.0730	$C_{14}H_9NO_2$ $C_{14}H_7NO_2$ $C_{14}H_7NO$ $C_{13}H_9NO$ $C_{12}H_9N$	-0.5 2.0 -1.2 0.6 5.7	L4		(Ahmed and Chiron, 2014; Brezina et al., 2017; Hübner et al., 2014; Jelic et

											al., 2013; Koba et al., 2016; Li et al., 2013; Zhu et al., 2016)
TP 269A		$C_{15}H_{12}N_2O_3$	269.0920	3.8	12.4	251.0815 208.0757 180.0808	$C_{15}H_{10}N_2O_2$ $C_{14}H_9NO$ $C_{13}H_9N$	-0.4 -4.8 -3.8	L3	MS/MS spectra reported	(Ahmed and Chiron, 2014; Hübner et al., 2014; Jelic et al., 2013)
TP 269B		$C_{15}H_{12}N_2O_3$	269.0920	2.1	15.1	No MS/MS			L4		
TP 269C		$C_{15}H_{12}N_2O_3$	269.0920	0.8	16.8	251.0815 208.0757 180.0808	$C_{15}H_{10}N_2O_2$ $C_{14}H_9NO$ $C_{13}H_9N$	0.8 -0.4 2.4	L3	MS/MS spectra reported	(Ahmed and Chiron, 2014; Hübner et al., 2014; Jelic et al., 2013)
TP 269D		$C_{15}H_{12}N_2O_3$	269.0920	-2.5	17.1	251.0815 208.0757 196.0757 180.0808	$C_{15}H_{10}N_2O_2$ $C_{14}H_9NO$ $C_{13}H_9NO$ $C_{13}H_9N$	-0.8 -5.7 -7.1 4.6	L3	MS/MS spectra reported	(Ahmed and Chiron, 2014; Hübner et al., 2014; Jelic et al., 2013)

TP 269E		$C_{15}H_{12}N_2O_3$	269.0920	1	17.5	251.0815 226.0863 208.0757 180.0808	$C_{15}H_{10}N_2O_2$ $C_{14}H_{11}NO_2$ $C_{14}H_9NO$ $C_{13}H_9N$	-9.2 4.2 -2.4 -5.4	L3	MS/MS spectra reported	(Ahmed and Chiron, 2014; Hübner et al., 2014; Jelic et al., 2013)
TP 271A/B		$C_{15}H_{14}N_2O_3$	271.1077	-1.3	14.0	253.0971 236.0706 210.0913 208.0757 180.0808	$C_{15}H_{12}N_2O_2$ $C_{15}H_9NO_2$ $C_{14}H_{11}NO$ $C_{14}H_9NO$ $C_{13}H_9N$	1.8 -5.1 2.7 1.0 -0.4	L3	MS/MS spectra reported	(Hübner et al., 2014; Jelic et al., 2013; Li et al., 2013)
		$C_{15}H_{14}N_2O_3$	271.1077	-3.54	14.8	236.0706 210.0913 180.0808	$C_{15}H_9NO_2$ $C_{14}H_{11}NO$ $C_{13}H_9N$	-6.4 1.2 1.7	L3	MS/MS spectra reported	(Hübner et al., 2014; Jelic et al., 2013)
TP 285A		$C_{15}H_{12}N_2O_4$	285.0867	0.2	7.4	267.0764 249.0659 239.0815 221.0709 212.0706	$C_{15}H_{10}N_2O_3$ $C_{15}H_8N_2O_2$ $C_{14}H_{10}N_2O_2$ $C_{14}H_8N_2O$ $C_{13}H_9NO_2$	5.0 -7.4 0.4 -0.2 -7.9	L3	MS/MS spectra reported	(Ahmed and Chiron, 2014)
TP 285B		$C_{15}H_{12}N_2O_4$	285.0867	-0.5	13.2	267.0764 239.0815 193.0760	$C_{15}H_{10}N_2O_3$ $C_{14}H_{10}N_2O_2$ $C_{13}H_8N_2$	-4.9 -5 -3.8	L3	MS/MS spectra reported	(Ahmed and Chiron, 2014)

TP 287		$C_{15}H_{14}N_2O_4$	287.1026	0.6	15.6	236.0706 223.0866 210.0913 180.0808	$C_{15}H_9NO_2$ $C_{14}H_{10}N_2O$ $C_{14}H_{11}NO$ $C_{13}H_9N$	4.2 0.9 -4 0.7	L3	Elucidation

667

668 **Table 2.** List of the CBZ TPs detected per commodity during the vegetable growth ("X"
669 indicates presence and "-" absence of the TPs in the studied matrices).

Compound	Irrigation conditions				
	SW[a]+CBZ[b]		SW+CBZ+UVC[c]		
	Lettuce	Peat	Water	Lettuce	Peat
CBZ	X	X	X	X	X
ACRI	X	X	X	X	X
TP 194	-	X	X	-	X
ACRO	X	X	X	X	X
TP 208A	-	-	X	-	-
TP 208B	-	-	X	-	-
TP 224A	X	X	X	X	X
TP 224B	-	X	X	-	X
TP 239	X	X	X	X	X
TP 253A	-	-	X	-	-
TP 253B	X	X	X	X	X
EPOX	X	X	X	X	X
OX	-	-	X	-	-
TP 267A	-	-	X	-	-
TP 267B	-	-	X	-	-
TP 269A	-	-	X	-	-
TP 269B	-	-	X	-	-
TP 269C	X	-	X	X	-
TP 269D	-	-	X	-	-
TP 269E	-	-	X	-	-
TP 271A	X	-	X	X	-
TP 271B	X	X	X	X	X
TP 285A	-	-	-	-	X
TP 285B	-	X	-	-	X
TP 287	-	-	X	-	-

670 [a]SW,Synthetic water; [b]CBZ, Carbamzepine; [c]UVC, Ultraviolet-C treatment
671

672 **Table 3**. Concentrations (ng g^{-1}) of CBZ and the validated TPs found in peat and lettuce
673 samples in both irrigation experiments.

Week of plant growth	ACRI	ACRO	EPOX	CBZ
	Peat[a] / Lettuce[b] irrigated with SW[c]+CBZ[d]			
2	ND[e] (-) / ND (-)	28 ± 1.2 / ND (-)	15 ± 1.9 / 34 ± 3.6	1844 ± 154 / 658 ± 26
4	4.4 ± 1 / ND (-)	23 ± 6.7 / ND (-)	40 ± 2.7 / 85 ± 7.5	1795 ± 598 / 803 ± 42
6	2.8 ± 0.8 / ND (-)	23 ± 3.1 / 1.3 ± 0.26	43 ± 10 / 82 ± 4.7	2264 ± 357 / 1112 ± 23
8	3.2 ± 1.2 / ND (-)	27 ± 10 / 1.7 ± 0.69	59 ± 4.4 / 103 ± 3.9	2260 ± 308 / 1090 ± 27
10	9.8 ± 1.1 / 0.65 ± 0.1	28 ± 3.3 / 5.1 ± 1.2	85 ± 6.7 / 187 ± 6.1	3097 ± 377 / 1749 ± 49
	Peat / Lettuce irrigated with SW+CBZ+UVC[f]			
Week of plant growth	ACRI	ACRO	EPOX	CBZ
2	13 ± 1.7 / ND (-)	42 ± 1.3 / ND (-)	23 ± 0.94 / 24 ± 1.1	1000 ± 13 / 419 ± 9
4	8.3 ± 5.5 / ND (-)	36 ± 6.7 / ND (-)	36 ± 1.8 / 37 ± 1.6	1381 ± 42 / 556 ± 16
6	15 ± 4.9 / ND (-)	39 ± 3.1 / 0.76 ± 0.23	51 ± 2.4 / 57 ± 3.5	1655 ± 16 / 828 ± 68
8	21 ± 2.3 / ND (-)	39 ± 10 / 1.9 ± 0.26	66 ± 1.6 / 79 ± 6.1	1945 ± 76 / 889 ± 69
10	23 ± 1.4 / 0.33 ± 0.05	44 ± 3.3 / 2.5 ± 0.56	92 ± 2.9 / 100 ± 11	2265 ± 22 / 1018 ± 100

674 [a]Peat concentrations in dry weight, d.w.; [b]Lettuce concentrations in wet weight, w.w.; [c]Synthetic water;
675 [d]Carbamazepine; [e]Not Detected; [f]Ultraviolet-C treatment
676
677

Figure captions

Figure 1. Extracted ion chromatogram (XIC) and MS/MS spectra of EPOX, OX and TP 253B from a UVC treated irrigation water sample. Comparison of the MS/MS spectra of EPOX and OX with the analytical standard.

Figure 2. Evolution on the abundances of the CBZ TPs detected in lettuce and peat samples during the plant growth.